Clostridium difficile Infection (CDI) Prevention Primer

Last reviewed 03/04/16. Disclaimer: The findings and conclusions in this presentation are those of the authors and do not necessarily represent the official position of the Centers for Disease Control and Prevention.



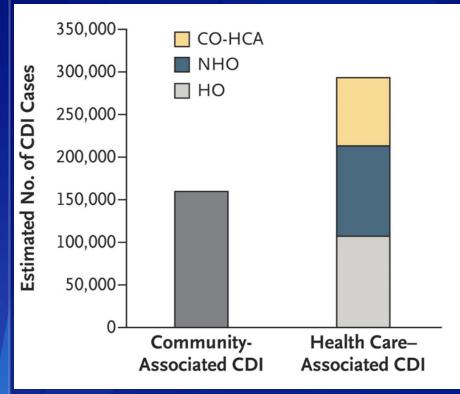
Outline

Background on *Clostridium difficile* Infection (CDI)

- Burden
- Pathogenesis
- Epidemiology
- General Infrastructure for CDI Prevention
- Basic Practices and Special Approaches for CDI Prevention
- Implementing a Practical Approach to CDI Prevention
- Resources and References

Background: Burden, Pathogenesis, Epidemiology

Estimated Annual U.S. Burden



Estimated U.S. Burden of CDI, According to the Location of Stool Collection and Inpatient Health Care Exposure, 2011.

CO-HCA: Community onset healthcare-associated **NHO:** Nursing home onset **HO:** Hospital onset

453,000 CDI cases¹

- 293,000 healthcare-associated
 - 107,000 hospital-onset
 - 104,000 nursing home-onset
 - 81,000 community-onset, healthcarefacility associated
- 160,000 community-associated
 - 82% associated with outpatient healthcare exposure

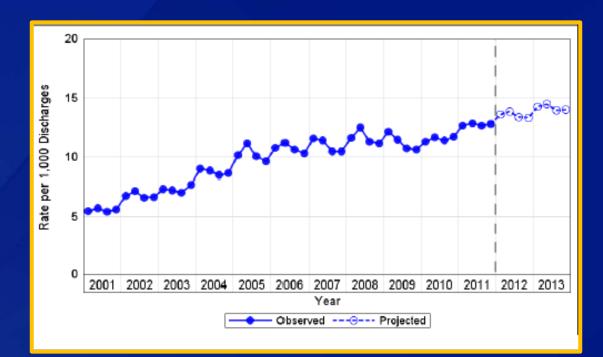
Overall, 94% of CDI cases related to healthcare

- 29,000 deaths
- \$4.8 billion in excess healthcare costs²

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Healthcare Burden

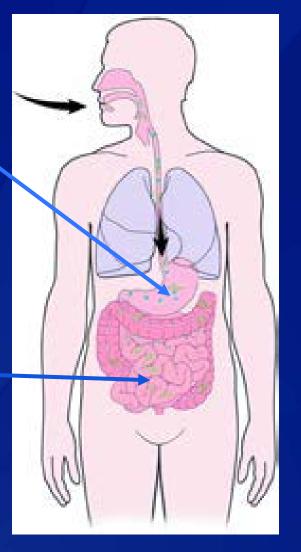
- *C. difficile* most commonly reported pathogen in 2011 multistate prevalence survey of healthcare-associated infections (HAI)¹
 - 12.1% of 452 HAIs caused by CDI
 - Rates of CDI per 1,000 discharges have risen through 2013²



Pathogenesis of CDI

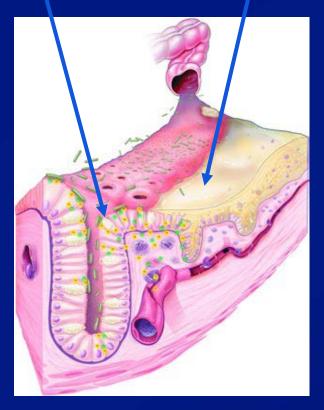
1. CDI spores survive in the environment for long periods of time. Following ingestion, they traverse the acidic environment of the stomach.

2. Spores germinate within the intestine.



3. Altered lower intestine flora (due to antimicrobial use) allows proliferation of *C. difficile* in colon.

> 4. Toxin A & B Production leads to colon damage +/- pseudomembrane.



Epidemiology: Epidemic Strain

- BI/NAP1/027, toxinotype III
- First emerged in 2000¹
- Associated with healthcare²
- More resistant to fluoroquinolones³
- Greater virulence
 - Associated with more severe disease and mortality⁴
 - Increased toxin A and B production^{4,5}
 - Polymorphisms in binding domain of toxin B⁵

Epidemiology: Host Factors



Advanced age

- Incidence higher among females, whites, and persons > 65 years¹
- Death more common in persons > 65 years (5x greater risk)²



Underlying illness and medical history

- 79% of 7421 patients with CDI had a comorbid condition²
- 38% of 585 patients with NAP1 strain had ED visit in previous 12 weeks²
- Tube feeds³

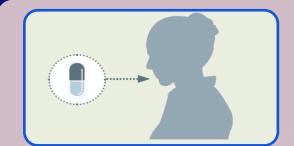


Immunosuppression

- Inflammatory bowel disease²
- Immune-suppressive treatment²
- Hematological malignancy/stem cell transplant (15-25x greater risk)⁴

1. Lessa et al. N Engl J Med 2015; 372(9):825-834. 2. See et al. Clin Infect Dis 2014; 58(10):1394-1400. 3. Bliss et al. Ann Intern Med 1998; 129:1012-1019.
4. Kombuj et al. Infect Control Hosp Epidemiol 2016; 37:8-15.

Epidemiology: Modifiable Risk Factors



Exposure to antibiotics

High Risk:

- Fluoroquinolones¹
- 3rd and 4th generation cephalosporins, clindamycin, carbapenems²





Exposure to *C. difficile* **spores**

- Spores can remain viable for months³
- Contamination is increased in rooms of patients with active CDI ^{4,5}
- Hands of patients and personnel are easily contaminated⁵

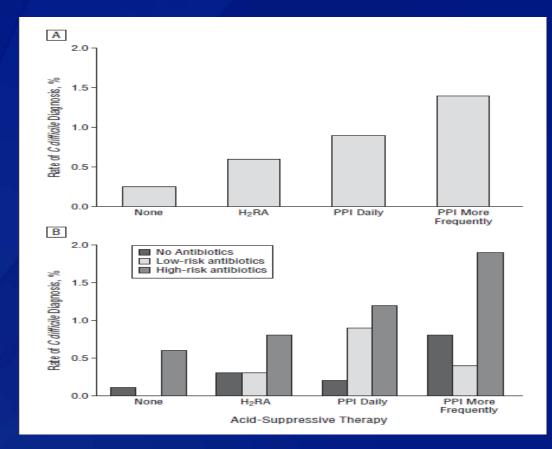
Gastric acid suppression

- Data, though inconsistent, implicate proton pump inhibitor (PPI) use^{1,4,6,7}
- More study is needed to link restriction of PPI use with decreased CDI incidence⁸

- 1. Pepin et al. Clin Infect Dis 2005; 41(9):1254–1260.
- 2. Hensgens et al. J Antimicrob Chemother 2012; 67(3):742-748.
- 3. Weber & Rutala. Infect Control Hosp Epidemiol 2011; 32: 207-209.
- 4. Dubberke et al. Am J Infect Control 2007; 35:315-318.

- 5. Shaugnessy et al. Infect Control Hosp Epidemiol 2011; 32;201-206.
- Linney et al. Can J Hosp Pharm 2010; 63(1):31–37.
- 7. Buendgens et al. J Crit Care 2014; 696:e11-15.
- 8. Dubberke et al. Infect Control Hosp Epidemiol 2014; 35(6):628-645.

Gastric Acid Suppression and CDI Risk *Dose Response and Interactions with Antibiotics*



Use of Proton Pump Inhibitors (PPI) and CDI

February 8, 2012 – FDA warning on PPI Use



U.S. Food and Drug Administration Protecting and Promoting *Your* Health

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Drugs

Home

Home Drugs Drug Safety and Availability

Drug Safety and Availability	
Drug Alerts and Statements	
Importing Prescription Drugs	

FDA Drug Safety Communication: *Clostridium difficile*-associated diarrhea can be associated with stomach acid drugs known as proton pump inhibitors (PPIs)

Defining Outbreaks and Hyperendemic CDI

• What is an outbreak?

- Increase in CDI that is greater than expected by chance alone
- Can be facility-wide, unit specific, or occurring within the community

What is hyperendemicity?

- Persistently high rates of CDI compared to past rates or compared to similar facilities/units
- Example: Excess infections above a prevention goal as indicated by the Cumulative Attributable Difference (CAD) metric in an NHSN Targeted Assessment for Prevention (TAP) report

Data for Action Using the CDC Targeted Assessment for Prevention (TAP) Strategy

Target	Assess	
 Generate TAP Reports using NHSN Identify locations with excess HAIs 	• Assess targeted locations for potential gaps in infection control	 Use tailored prevention strategies to address identified gaps
• Engage targeted locations in focused prevention efforts		

An elevated CDI CAD on a hospital's TAP report can identify the need to initiate an infection prevention assessment. In many cases, a specific unit or units in the hospital account for the majority of cases.

Target Units or Facilities with Excess Infections

Identify facilities or units with opportunities for improvement using a TAP Report generated in the National Healthcare Safety Network (NHSN).

TAP Strategy 'How To' Guide

for the <u>Individual Facility User</u> Targeted Assessment for Prevention: Using Data for Action <u>www.cdc.gov/hai/prevent/tap.html</u>

The Targeted Assessment for Prevention (TAP) Strategy is a framework for quality improvement that offers a focused approach to infection prevention for healthcare facilities, healthcare systems, public health, and quality improvement partners. This strategy can be used to identify facilities and units with a high burden of healthcare-associated infections (HAIs) so that specific gaps in infection prevention can be identified and addressed. The TAP strategy incorporates the TAP reports generated in CDC's National Healthcare Safety Network (<u>NHSN</u>) along with standardized assessment tools and accompanying implementation strategies.

TAP Strategy 'How To' Guide

for the <u>Group User</u> Targeted Assessment for Prevention: Using Data for Action www.cdc.gov/hai/prevent/tap.html

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Assess Infection Prevention Practices

Assess policies and practices related to CDI prevention

- Leadership
- Training, auditing/feedback
- Antibiotic stewardship
- Early detection and isolation
- Appropriate testing practices
- Contact Precautions/hand hygiene
- Environmental cleaning

н.	Antibiotic Stewardship for CDI Prevention	Response Choices	Comments (and/or "As Evidenced By")
1.	Does your facility review appropriateness of antibiotics prescribed for treatment of other conditions (e.g., UTI) for patients with new or recent CDI diagnosis?		
2.	Does your facility educate providers about the risk of CDI with antibiotics?	OYesONo OUnk	
3.	Does your facility educate patients/family members about the risk of CDI with antibiotics?	Yes No Unk	
Do	es your facility monitor the use of the following antibiotics that are high	-risk for CDI:	
4.	Fluoroquinolones	Yes No Unk	
5.	3 rd /4 th generation cephalosporins?	Yes No Unk	
Do	es your facility use strategies to reduce the use of the following antibiot	ics that are high-risk for CDI:	
6.	Fluoroquinolones	Yes No Unk	
7	3 rd /4 th generation cephalosporins?	Yes No OUnk	

Prevent Infections with Tailored Measures

II. Antibiotic Stewardship				
Example Resources	Websites			
Get Smart for Healthcare – Implementation Resources Resources to assist in the implementation of hospital antibiotic stewardship programs, including guidelines, assessment tools, conceptual models, and a sample inter-facility infection control transfer form, from CDC	http://www.cdc.gov/getsmart/healthcare/implementation.htm !			
Get Smart for Healthcare – Stewardship Program Examples Links to hospital stewardship programs at various hospitals, success stories, and an interactive collection of charts and maps summarizing national and subnational data on antimicrobial use and resistance (Resistance Map), from CDC	http://www.cdc.gov/getsmart/healthcare/programs.html			
Get Smart for Healthcare - Checklist for Core Elements of Hospital Antibiotic Stewardship Programs A checklist to evaluate hospital antibiotic stewardship programs, from CDC	Checklist for Core Elements of Hospital Antibiotic Stewardship Programs Get Smart for Healthcare CDC			
Antimicrobial Stewardship Toolkit Best practices from an Antimicrobial Stewardship Collaborative, from the Greater New York Hospital Association and the United Hospital Fund	http://www.shea- online.org/Portals/0/GNYHA Antimicrobial Stewardship Toolk it FINALv2%20Dec2011.pdf			
Toolkit for Reduction of Clostridium difficile Infections Through Antimicrobial Stewardship: Possible Methods for Evaluating Antibiotic Use Table of possible metrics to use for measurement and evaluation of antibiotic use, from the Agency for Healthcare Research and Quality (AHRQ)	http://www.ahrq.gov/professionals/quality-patient- safety/patient-safety- resources/resources/cdifftoolkit/cdiffl2tools2c.html			

III. Early Detection and Isolation, Appropriate Testing				
Example Resources	Websites			
Algorithms for Prevention and Management of <i>Clostridium difficile</i> Infections in Long-term Care Facilities	http://www.health.state.mn.us/divs/idepc/diseases/cdiff/hcp/l tcalgorithms.pdf			
Decision-making strategies for enhancing early recognition, testing, and isolation of patients with CDI in long- term care facilities, from the Minnesota Department of Public Health				
C. difficile Infection Change Package: Preventing C. difficile Transmission and Infection Compilation of tools, including algorithms for testing and diarrhea decision trees that align with appropriate isolation and testing guidelines (pgs. 12, 13), from the Health Research & Educational Trust (HRET), American Hospital Association (AHA), and Partnership for Patients	http://www.hret-hen.org/topics/cdi/13-14/2014- CDIChangePackage508.pdf			
Bristol Stool Form Scale Scale tool that provides an objective way to differentiate between various types of stool forms and recognize diarrhea, from the National Institutes of Health (NIH)	http://bowelcontrol.nih.gov/bristol.aspx			

General Infrastructure for CDI Prevention

Leadership

- Need to understand organizational culture and change it when it hinders performance¹
- Direct evidence linking leadership to infection rates is limited but consistent themes have been identified²

Leadership traits that may assist in preventing infections³:

Proactive, positive, visible Supportive of change Clear responsibilities Clear policies

Leadership traits that may be associated with risk²:

Reactive

Laissez faire management style

Failure to assign responsibility and maintain accountability

Wide span of control



Capacity may be affected by complex management issues

Low Nurse Staffing Ratios

Nurse to patient staffing ratios inversely associated with healthcare-associated infections (UTI and SSI).¹

High Occupancy Inpatient wards with occupancy rates of 80-90% had CDI rates 56% higher than during baseline occupancy rates (0-69%).²

Feeling Overwhelmed Stress and chaos associated with poorer infection prevention practices.³

Strong leadership at the unit level and above is likely a prerequisite for infection prevention improvements⁴

1. Cimiotti et al. Am J Infect Control 2012; 40:486-490.

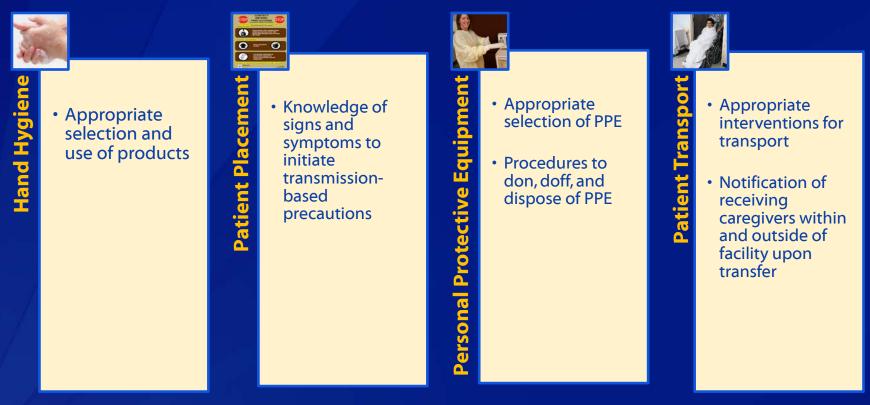
2. Ahyow et al. Infect Control Hosp Epidemiol 2013; 34:1062-1069.

Sinkowitz-Cochran et al. Am J Infect Control 2012; 40:138-143.
 Griffiths et al. J Hosp Infect 2009; 73:1-14.

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Competency

Healthcare personnel education and competencies are foundational to successful performance



Sustainable Quality Improvement

Participants in The Institute for Healthcare Improvement 100,000 Lives Campaign believed that data feedback, buy in, hardwiring (incorporation into daily activities), and leadership support were essential to sustainability.



General Infrastructure *Surveillance and Ongoing Measurement*

Process and outcome measures should be collected using ongoing, longitudinal methods with regular feedback to healthcare personnel.¹

Process Measures	 Prospective, actionable, useful for feedback about unit implementation of prevention measures² Examples include: Appropriate Isolation Hand hygiene and PPE adherence 		
Outcome Measures	 Useful to target assessments and prevention measures Examples include: Hospital-onset CDI Community-onset, healthcare facility-associated CDI 		

Basic Practices and Special Approaches for CDI Prevention

Overview

- <u>Basic practices</u> are prevention measures that should be in place at all times.
- Facilities should consider adopting some or all of the <u>special approaches</u> whenever ongoing opportunities for improvement are identified or as indicated by risk assessment.

Basic Practices	Special Approaches
Appropriate use of antimicrobials	Antimicrobial stewardship program
Hand hygiene per CDC/WHO recommendationsMeasure healthcare personnel adherence	Hand hygiene with soap and water after glove removal following care of CDI patientsIntensify measurement of adherence
Contact Precautions for CDI patients Measure healthcare personnel adherence 	Presumptive Contact Precautions while laboratory results are pending Prolonged duration of Contact Precautions Intensify measurement of adherence
Cleaning and disinfection of equipment and environment	Use of EPA-approved sporicidal disinfectant Assess adequacy of room cleaning
Laboratory-based alert system for immediate notification to IP and clinical personnel of newly diagnosed CDI patients	
CDI surveillance, analysis, and reporting	
Educate healthcare personnel, patients, and families	

Antimicrobial Stewardship

Antimicrobial Stewardship

Exposure to any antimicrobial is the single most important risk factor for *C. difficile* infection (CDI).

• Antibiotic exposure has lasting impact on the microbiome.

- Risk of CDI is elevated (7-10 fold) during and in the 3 months following antimicrobial therapy^{1,2}
- 85-90% of CDI occurs within 30 days of antimicrobial exposure¹
- Target high risk antibiotics for CDI prevention
 - Fluoroquinolones³
 - 3rd/4th generation cephalosporins, carbapenems²

Seven Core Elements of Antimicrobial Stewardship

1. Leadership Commitment

Dedicating necessary human, financial, technological resources

2. Accountability

Appointing a single leader (physician or pharmacist) responsible for program outcomes

3. Drug Expertise

A single dedicated pharmacist with responsibility to improve antibiotic use

4. Tracking

Monitoring antibiotic prescribing and resistance patterns

5. Reporting

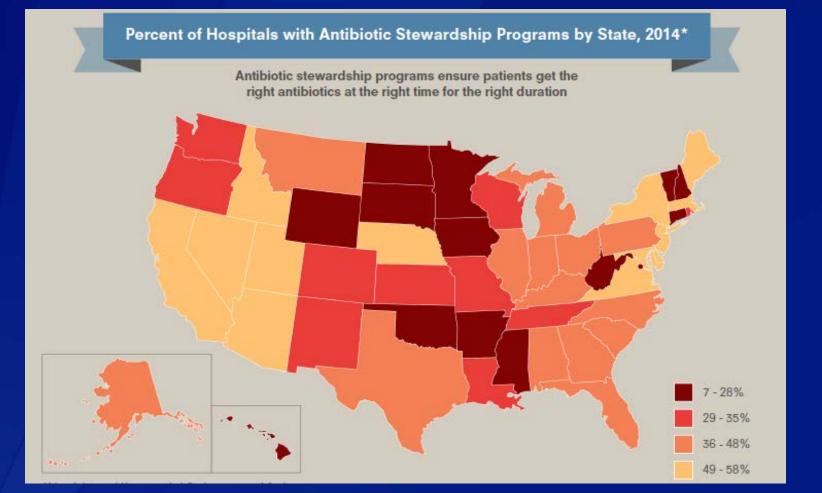
Feedback of information on antibiotic use and resistance to frontline providers

6. Education

Ongoing education of clinicians about resistance and optimal prescribing

7. Action

Implementing at least one recommended action



Currently 39% (1,642/4,184) of U.S. hospitals have an antibiotic stewardship program with all 7 core elements.

The national goal is 100% of hospitals by 2020.

Leaders Committed to Antibiotic Stewardship

Consider making your organization's commitment to Antibiotic Stewardship public on the CDC Website.

Examples:

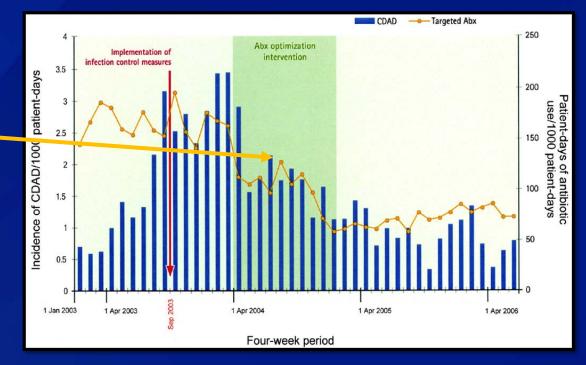
- Hospital Corporation of America has committed to continue current collaborations with CDC about this issue, develop and implement new clinical decision support and real-time antibiogram tracking to rapidly respond to lab results, catch bug-drug mismatches, implement strategy to prevent health-care associated infections in adult intensive care unit patients, and strengthen national efforts to identify and report cases of antibiotic resistance.
- **The Joint Commission** has committed to include stewardship as part of onsite surveys.

Full instructions on submitting a commitment are available at: http://www.cdc.gov/drugresistance/federal-engagement-in-ar/stewardship-commitment/index.html

Stewardship Approach: Feedback

Non-restrictive feedback resulted in statistically significant reductions in incident CDI.

Reductions in CDI attained through antimicrobial stewardship surpassed those attained through infection control measures.



Tertiary Hospital in Quebec, 2003-2006

Stewardship Approach: Restriction

Restricting the use of ceftriaxone was associated with reduced rates of CDI.

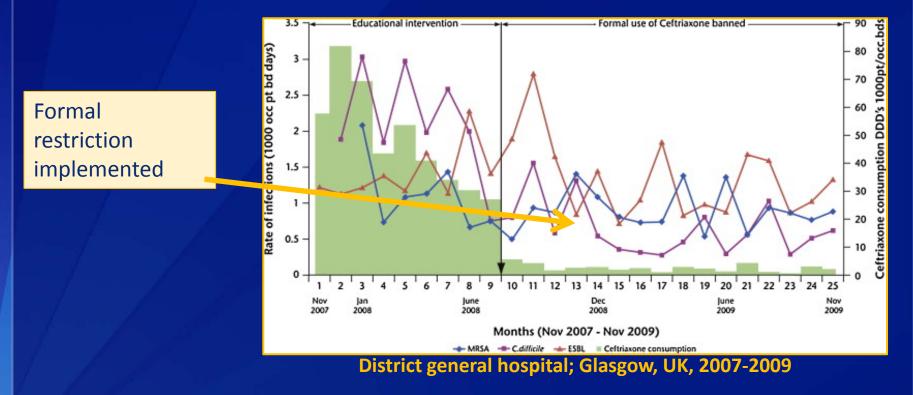


Fig. 1 Hospital-acquired methicillin-resistant *Staphylococcus aureus* (MRSA), *Clostridium difficile* and extended-spectrum β-lactamase (ESBL)-producing coliform rates following a restrictive antibiotic policy in a district general hospital over 2 years. pt/occ.bds, patient-occupied bed-days; DDDs, defined daily doses.

Early Detection and Isolation

Early Detection and Isolation

1) Focus testing on patients with clinically significant diarrhea,^{1,2} without other identified causes

- ≥ 3 liquid bowel movements (type 7) in 24 hours
- Stool conforms to shape of container

2) Utilize presumptive Contact Precautions* until infectious causes are ruled out

> The Bristol Stool Chart assists in objectively identifying stool characteristics²

LET'S TALK A NIDDK about Bowel Control Bristol Stool Form Scale Type Description Image Separate hard lumps, like nuts Type 1 Sausage-shaped but lumpy Type 2 Like a sausage or shake but with cracks on th Type 3 Type 4 Like a sausage or snake, smooth and soft Type 5 Soft blobs with clear-cut edges Type 6 Fluffy pieces with ragged edges, a mushy stoo Type 7 Watery, no solid pieces Taxt reproduced with permission from Lawlo SJ, Heaton KW, Boandinestan Joarna' of Gastroenterology, 1997, 5255,505-4 01997 Internet Heathcare.

*presumptive use of Contact Precautions is a special approach

Dubberke et al. Infect Control Hosp Epidemiol 2014; 35(6):628-645.

Peterson & Robicsek. Ann Intern Med 2009; 151(3):176-179.

Patient and Family Education

Educate

- Patient education at the time of isolation is a critical intervention to reduce stress and anxiety¹
- Patient educational background may influence the preferred mode of education²

Engage

 94 of 100 patients believed that information about their infection would help them make choices that resulted in better care²

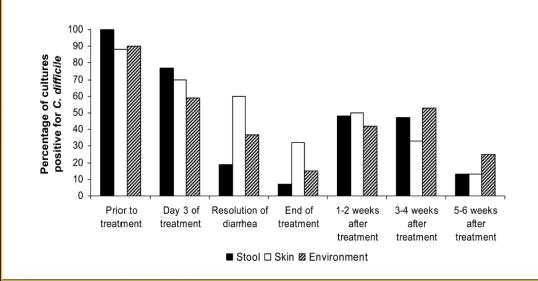
Empower

 Patients more likely to speak up to ensure adherence with hand hygiene if explicitly empowered to do so by healthcare personnel³

Contact Precautions and Hand Hygiene

Contact Precautions (CP)

- Contamination of the environment is highest prior to treatment¹
- Presumptive CP, while CDI test results are pending, may be used as a special approach whenever indicated by risk assessement²

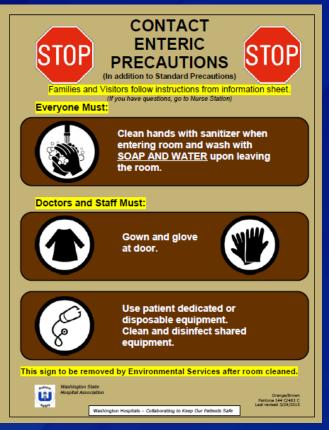


- Patients who have been treated may have asymptomatic shedding³
- **Prolonging the duration of CP** until discharge is a special approach based on evidence of continued shedding of spores after diarrhea resolves (especially up to 4 weeks after treatment ends)²

1. Bobulsky et al. Clin Infect Dis 2008; 46(3):447-450

^{2.} Dubberke et al. Infect Control Hosp Epidemiol 2014; 35(6):628-645

Contact Precautions



Isolation Precaution Signage: http://www.wsha.org/qualitysafety/projects/standardization/

- Don gown and gloves upon room entry¹
- Use disposable or dedicated patient care equipment^{1,2}
 - Electronic thermometers have been associated with transmission
- Communicate Contact Precautions^{2,3}
 - at room entrance
 - at handoff
 - during transfers

1. CDC Guideline for Isolation Precautions: <u>http://www.cdc.gov/hicpac/2007IP/2007ip_appendA.html</u> 2. Dubberke et al. Infect Control Hosp Epidemiol 2014; 35(6):628-645 3. Hsu et al. Am J Gastroenterol 2010; 105(11):2327–2339

Cohorting of CDI Patients

- Private rooms are preferred for patients with fecal incontinence¹
- Cohorting of patients^{1,2}
 - Cohort patients with same multidrug-resistant organisms only
 - Ensure healthcare personnel follow appropriate isolation practices when moving between patients
 - Perform hand hygiene when donning or doffing PPE
 - Gloves and gowns must be changed and HH performed between each patient

2. CDC Guideline for Isolation Precautions: http://www.cdc.gov/hicpac/2007IP/2007ip_appendA.htm

^{1.} Dubberke et al. Infect Control Hosp Epidemiol 2014; 35(6):628-645.

Elements of a Multi-Modal Program to Improve Hand Hygiene

A robust, ongoing hand hygiene program should be in place

Program Component	Action Item
Ongoing monitoring of HAI incidence	Setting specific feedback about incident CDI
Administrative support and leadership	Clear policies and messages about hand hygiene
Multidisciplinary team	Members of administration, clinical leaders, and front line staff collaboration to increase adherence
Ensure accessibility of supplies	Point of care hand hygiene should be within arms reach of healthcare personnel
Reinforce behavior and accountability	Contests, incentives, recognition tickets, notice letters, personnel action
Provision of reminders	Posters, just-in-time training, inclusion in checklists
Ongoing monitoring of adherence and feedback on compliance	Direct observation, product use monitoring, real- time feedback, monthly posting on adherence,

Hand Hygiene and Care of the Patient with CDI



- Hand hygiene policies should promote preferential use of alcohol-based hand rub (ABHR) over soap and water in all clinical situations except when hands are visibly soiled (e.g., blood, body fluids) or after caring for a patient with known or suspected *C. difficile* or norovirus during an outbreak or if endemic rates are high.
- Strict adherence to glove use is the most effective means of preventing hand contamination with *C. difficile* spores¹
 - Spores not killed by ABHR and may be difficult to remove even with thorough hand washing ^{2,3}
 - Although there have been no studies demonstrating a decrease in CDI infection with use of soap and water as opposed to ABHR⁴, because ABHR is not effective against spores, facilities may consider routine use of soap and water after glove removal during care of patients with *C. difficile* infection even absent an outbreak.
- Measuring compliance is a basic practice and critical to success
 - Ensure technique is good and re-educate if lapses identified

Soap and Water vs. Alcohol Hand Rub

Intervention	Mean log reduction (95% CI),			
Intervention 1	Intervention 2	log ₁₀ CFU/mL		
Warm water and plain soap	No hand hygiene	2.14 (1.74-2.54)		
Warm water and plain soap	Alcohol-based handrub	2.08 (1.69-2.47)		
Cold water and plain soap	No hand hygiene	1.88 (1.48-2.28)		
Cold water and plain soap	Alcohol-based handrub	1.82 (1.43-2.22)		
Warm water and plain soap	Antiseptic hand wipe	1.57 (1.18-1.96)		
Warm water and antibacterial soap	No hand hygiene	1.51 (1.12–1.91)		
Warm water and antibacterial soap	Alcohol-based handrub	1.46 (1.06-1.85)		
Cold water and plain soap	Antiseptic hand wipe	1.31 (0.92-1.71)		
Warm water and antibacterial soap	Antiseptic hand wipe	0.94 (0.55-1.34)		
Warm water and plain soap	Warm water and antibacterial soap	0.63 (0.23-1.02)		
Antiseptic hand wipe	No hand hygiene	0.57 (0.17-0.96)		
Antiseptic hand wipe	Alcohol-based handrub	0.51 (0.12-0.91)		
Cold water and plain soap	Warm water and antibacterial soap	0.37 (-0.03 to 0.76)		
Warm water and plain soap	Cold water and plain soap	$0.26 \ (-0.14 \ \text{to} \ 0.66)$		
Alcohol-based handrub	No hand hygiene	0.06 (-0.34 to 0.45)		

ABHR and CDI Rates

Figure 1. Use of alcohol hand rub by healthcare workers; Liters per 1,000 patient-days, per quarter 2000-2003

Data from 2000-2003 show no association between alcohol-based hand rub (ABHR) use and increase in CDI rates

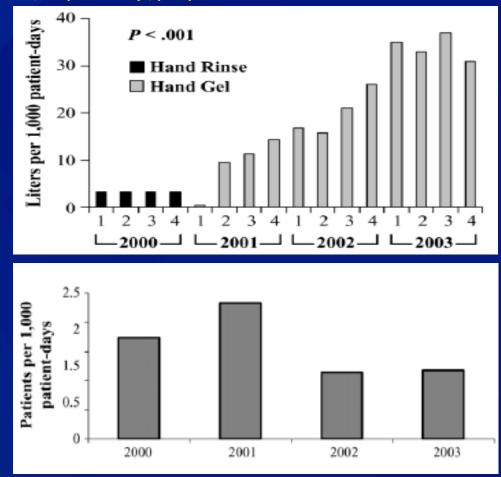


Figure 2. Number of patients with 1 or more tests positive for *C. difficile* toxin per 1,000 patient-days, 2000-2003

Importance of Gloves

Since spores may be difficult to remove from hands even with hand washing, <u>strict adherence to glove use</u>, and Contact Precautions in general, should be emphasized for preventing *C. difficile* transmission via the hands of healthcare personnel.

Importance of Gloving

- Environmental contamination may be increased during outbreaks and in locations with hyperendemicity¹
- Glove use is most effective means of preventing contamination of the hands of HCPs with *C. difficile* spores from symptomatic patients
- Universal gloving is a special approach for use when indicated by risk assessment²

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Remember Patient Hands!

A randomized trial of soap and water wash versus alcohol hand rub shows soap and water are more effective at removal of *C. difficile* spores on patient hands.

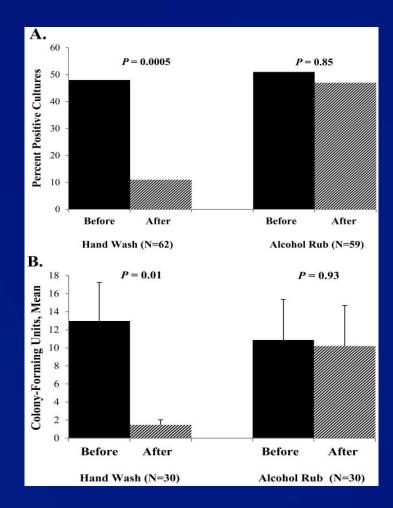


Figure 1: Percent of patient hands with positive *Clostridium difficile* cultures.

Environmental Cleaning and Disinfection

Key Principles

- There is no substitute for meticulous cleaning^{1,2,3}
- Cleaning reduces spores in the environment²
- Disinfectants with a sporicidal claim inactivate spores²
- Monitoring and feedback optimize performance¹
- Policies should clearly define who is responsible for cleaning and disinfecting environmental surfaces and equipment⁴

Cleaning	The removal of organic debris using vigorous wiping and or scrubbing until all visible soil is removed ¹
Disinfection	Removal or inactivation of some or all pathogens on inanimate objects ¹

1. Carling Am J Infect Control 2013; 41:S20-25.

2. Rutala et al. Infect Control Hosp Epidemiol 2012; 33(12):1255-1258.

3. CDC Guidelines for Environmental Control in Healthcare Facilities: http://www.cdc.gov/hicpac/pdf/guidelines/eic_in_HCF_03.pdf

4. CDC Guidelines for Disinfection and Sterilization in Healthcare Facilities: http://www.cdc.gov/hicpac/pdf/guidelines/Disinfection_Nov_2008.pdf

Mitigating Risks Through Environmental Cleaning and Disinfection



<mark>gh touch items</mark>

Facilities must ensure adequate cleaning of^{1,2} high touch surfaces in the patient environment.

- Examples:
- Bed rails
- Restroom hand rails
- Bed side tables
- See CDC checklist for full list*



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Disinfectant

- Facilities must ensure proper preparation and use of disinfectants^{3.} Including appropriate:
 Dilution
 - Storage
 - Application
 - Contact Time



Proper use of disinfectants

- Factors that influence disinfectant effectiveness³:
 - Porosity of surface
- Crevices or ridges
- Facilities must ensure cross contamination is avoided

1. Dubberke et al. Infect Control Hosp Epidemiol 2014; 35(6):628-645.

2. CDC Options for Evaluating Environmental Cleaning: <u>http://www.cdc.gov/hai/toolkits/Evaluating-Environmental-Cleaning.html</u>

3. CDC Guidelines for Environmental Control in Healthcare Facilities: http://www.cdc.gov/hicpac/pdf/guidelines/eic_in_HCF_03.pdf

Environmental Contamination

- Patients with asymptomatic carriage may be an important source of environmental contamination¹
- Patients that have been treated may continue to shed spores²

In a cross sectional survey of 6 facilities³:

- NAP1 strain recovered at each facility
- All rooms housing a CDI patient yielded positive cultures for *C. difficile*
- 4/12 (33%) of rooms without a CDI patient yielded positive cultures for *C. difficile*

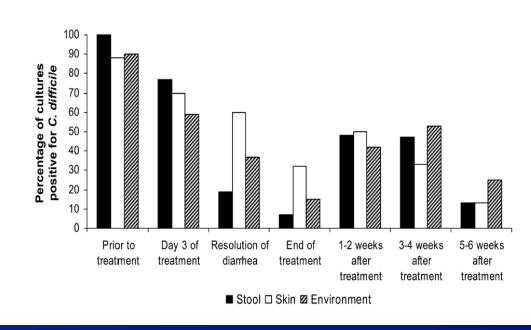


Figure from Sethi, 2010

2. Sethi et al. Infect Control Hosp Epidemiol 2010; 31:21-27.

Spore Removal

- Meticulous cleaning with any cleaner/disinfectant reduces the number of spores in the environment¹
- However, greater reduction and inactivation of spores is achieved when a sporicidal agent is used¹
- Removal of spores influenced by contact time (duration of wetness) and texture of surface being cleaned²

Technique	Reduction in Spores	Dry Time
Wiping with any disinfectant	> 2.9 log ₁₀	2-6 minutes
Spraying (no wipe) with sporicide	3.4 log ₁₀	28-40 minutes
Wiping with sporicide	3.9 log ₁₀	2-6 minutes

EPA-Approved Disinfectants Effective Against *C. difficile*

- Before-after intervention studies demonstrated benefit of sporicidal disinfectants in units with high endemic CDI rates¹
- Environmental contamination indirectly increases risk of cross contamination, likely via the hands of healthcare personnel²

When using sporicidal disinfectants¹:

- Avoid toxicity to patients and environmental services staff
- Avoid damage to equipment
- Ensure method to communicate when sporicidal disinfectants should be used

A current list of EPA-approved disinfectants with sporicidal claim is available at: <u>http://www.epa.gov/pesticide-registration/list-k-epas-registered-antimicrobial-products-</u> <u>effective-against-clostridium</u>

Inactivation of *C. difficile* Spores

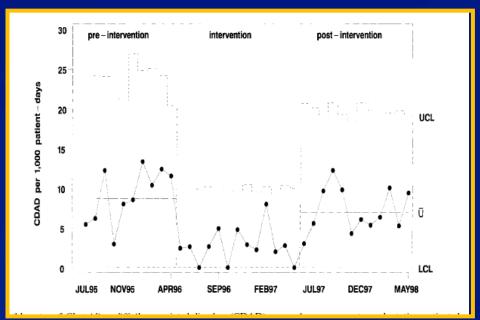
Sporicidal disinfectants most effective in reducing CDI when endemic rates are high.

Bone Marrow Transplant Unit CDI Rates/ 1000 pt. days:

- Pre intervention: 8.6
- Intervention: 3.3
- Post-intervention: 8.1

No reduction:

- Neurosurgical ICU: 3.0
- General medical ward: 1.3

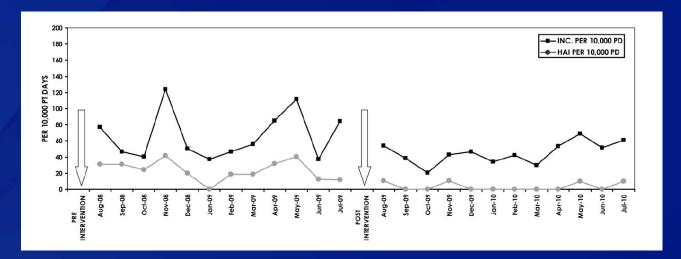


CDI rates in bone marrow transplant unit

Reducing CDI Using a Sporicidal Wipe for Cleaning

- Before/after study in two high-risk medical wards
- Intervention:
 - Daily and terminal cleaning of all rooms with ATP monitoring before/after (similar pass rate)
 - Quaternary ammonium compound <u>before</u>
 - Hypochlorite wipes with 10 minute contact time after

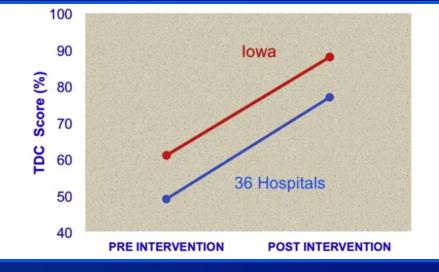
• Results: 24.2 to 3.6 cases per 10,000 patient-days (85% decline)



Evaluating Cleanliness

Objective assessment of cleaning assists in optimizing performance^{1,2}

- Monitoring of surfaces using florescent gel resulted in significant improvements in cleaning
- Educational interventions with environmental services staff resulted in sustained improvement



Terminal Disinfection and Cleaning

1. Carling. Am J Infect Control 2013; 41:S21-25.

2. CDC Options for Evaluating Environmental Cleaning: http://www.cdc.gov/hai/toolkits/Evaluating-Environmental-Cleaning.html

Methods of Evaluating Cleanliness

Evaluating Patient Zone Environmental Cleaning

Method	Ease of Use	Identifies Pathogens	Accuracy	Useful for Teaching	Use in Programmatic Monitoring
Direct observation	Low	No	Variable	Yes	Difficult
Culture swab	High	Yes	High	No	No
Agar culture system	Moderate	Possible	Moderate	No	Possible*
Fluorescent system	High	No	High	Yes	Yes
ATP Bioluminescence	High	No	Variable	Yes	Possible*

* Measures cleanliness at that moment but NOT the process of cleaning

Using ATP Bioluminescence To Evaluate Cleanliness

Facilities using Adenosine Triphosphate (ATP) bioluminescence to monitor cleaning should carefully consider the range and diversity of results¹

Researchers, using a convenience sample of 500 surfaces, determined²:

- 378 (76%) surfaces had fluorescent marker removed
- 225 (45%) had ATP bioluminescence measurement indicating cleanliness
- 384 (77%) had aerobic colony counts indicating cleanliness

Method	Pro	Con
Fluorescent Gel	Easy to apply – Stable in Environment Removal accurately signifies cleaning ³ Evaluates cleaning process ³ Effective training tool for ES staff ³	Requires application before cleaning and reassessment after cleaning
ATP Bioluminescence	Ease of use Evaluates current cleanliness ³ Effective training tool for ES staff	Does not evaluate cleaning process ³ ATP failures may not correlate with fluorescent gel or CFU counts ^{3,4} Disinfectants may confound results ⁴ Facility and surface specific benchmarks may be needed ^{3,4,5} Specificity ~ 57% ⁴
		vey et al. J Hosp Infect 2011; 77:25-30. ma & Malik Int J Hyg Env 2013; 216:115-125.
Carling Am J Infect Conti		

Ultraviolet Light and Hydrogen Peroxide Fogging

- Data currently insufficient to recommend inclusion of these methods in a CDI prevention program¹
- Standard room cleaning and disinfection found to be suboptimal when UV disinfection was used²
 - Consistent cleaning attributed to 2 interventions:
 - Dedicated team
 - Supervisory housekeeping staff or IP staff checked rooms post cleaning



Appropriate Testing and Laboratory Practices

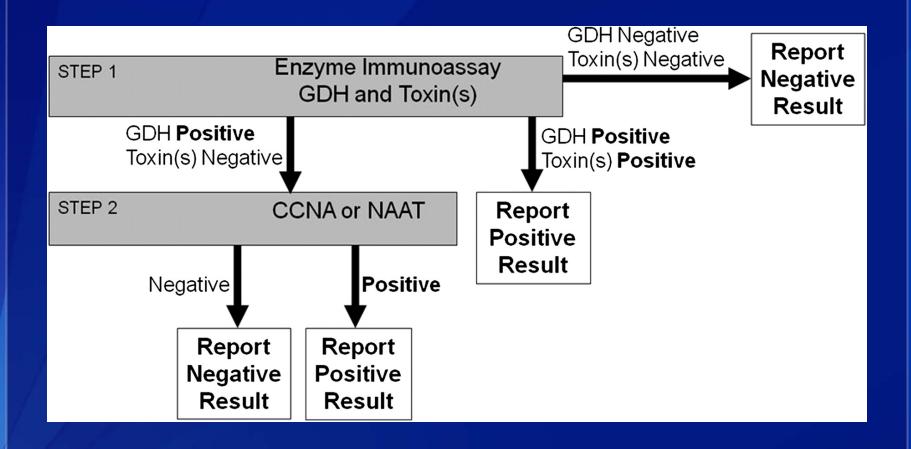
Guidance from the American Society for Microbiology

- Toxin A/B enzyme immunoassays have low sensitivity and should not be used as stand alone tests.¹
- Highly sensitive screening tests like glutamate dehydrogenase antigen assays (GDH) should have positive results confirmed.
- Nucleic acid amplification that detects *C. difficile* toxin genes may be used as a stand alone test.
- Repeat testing, testing of formed stool, and testing for cure should be avoided.²

Regardless of testing method, ensure appropriate testing to optimizing test performance!

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Typical Diagnostic Algorithm for Detection of Toxigenic *C. difficile* in Stool



Understanding Predictive Value

True prevalence in the population tested for *C. difficile* greatly impacts predictive value of diagnostic tests

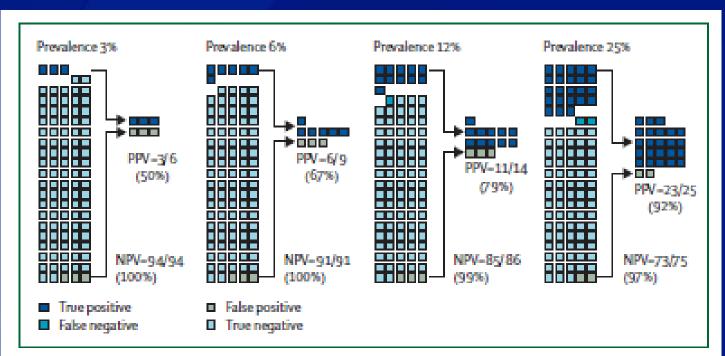


Figure 1: Effect of varying prevalence on the PPV and NPV of a theoretical Costridium difficile toxin assay with a sensitivity of 92% and a specificity of 97%

NPV-negative predictive value. PPV-positive predictive value.

Repeat Testing

 Testing with a low sensitivity test and repeat testing are not recommended¹

- Results in increased false positive results
- Pseudo-outbreaks related to false positives and repeated testing have been identified^{2,3}

Table 2.	Table 2. Model of Results for Toxigenic Clostridium difficile Detection When Testing Is Repeated*											
Test Sequence	EIA				qPCR							
Sequence	Tested, n	True Positive, n	PPV	False Positive, n	Undetected Disease, n	Remaining Negative Results, <i>n</i>	Tested, n	True Positive, n	PPV	False Positive, <i>n</i>	Undetected Disease, n	Remaining Negative Results, <i>n</i>
First	1000	73	0.75	24	27	903	1000	93	0.78	26	7	881
Second	903	18	0.45	22	9	863	881	7	0.23	23	0	851
Third	863	7	0.25	21	2	835	-	-		-	-	
Fourth	835	1	0.05	20	1	814	-	-		-	-	
Fifth	814	1	0.05	20	0	793	-	-		-	-	
Total		100†		107				100‡		49		

EIA = enzyme immunoassay; PPV = positive predictive value; qPCR = quantitative real-time polymerase chain reaction.

* In this model, there are 1000 tested participants and *C. difficile* prevalence in the test population is 10%. Patients with negative results have tests repeated sufficiently to ensure that all true-positive results are captured. Assumptions for EIA: sensitivity = 73.3%; specificity = 97.6%; and test performance does not change when repeated (6). Assumptions for qPCR: sensitivity = 93.3%; specificity = 97.4%; and test performance does not change when repeated (6). \dagger Overall PPV = 0.48.

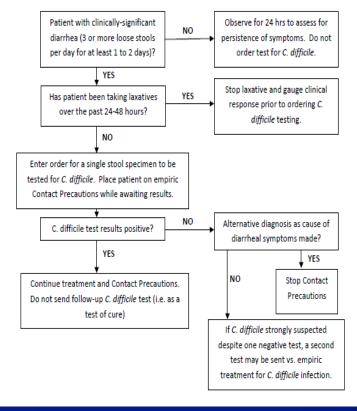
 \ddagger Overall PPV = 0.67.

Dubberke et al. Infect Control Hosp Epidemiol 2014; 35(6):628-645.
 Litvin et al. Infect Control Hosp Epidemiol 2009; 30:1166-1171.

Guidance for Appropriate Testing

 Tools that guide clinical decision making, such as algorithms, can assist in ensuring appropriate testing





Example of a CDI decision tool from Vanderbilt University Medical Center

1. Dubberke et al. Infect Control Hosp Epidemiol 2014; 35(6):628-645.

NHSN CDI Risk Adjusted SIR Accounts for More Sensitive Testing

Variables from Final Model to be included for Risk Adjustment in SIR Calculation

Factor	Description		
Intercept			
Facility Bed Size	> 245		
	101-245		
	≤ 100		
Teaching Type	Major		
	Graduate		
	Limited & Non		
CDI Test Type	NAAT (PCR)		
	EIA		
	All Other		
Prevalence	Continuous (no CO-HCFA)		

Data Sources and Submission

• CDI test type, facility bed size, and teaching type are collected on the required Annual Facility Survey

Implementing a Practical Approach to CDI Prevention

Targeted Assessment for Prevention (TAP) Strategy

Target facilities/units

Assess gaps in infection prevention in targeted areas **Prevent** infections by implementing interventions to address the gaps

CDI TAP reports available in NHSN

Work ongoing to use CO-CDI and other data sources to identify CDI cases from nursing homes and other community sources

www.cdc.gov/hai/prevent/tap.html

CDI Risk Assessment

- Conduct a risk assessment annually and whenever CDI goals are not met¹
 - Use CDC's Targeted Assessment for Prevention (TAP) Strategy
 - Pairing the results of a CDI facility assessment with the CDI Implementation Tool allows facilities to implement infection prevention strategies that most directly meet their needs.



CDI Case Review

- Review each CDI case (e.g., root cause analysis)
- Examine temporal and spatial relationships between cases to determine units at risk for transmission due to community-onset or hospital-onset CDI cases.

The CDI Implementation Tool includes a template for CDI Case Review

FEEDBACK

NHSN Resources for MDRO and C. difficile Infection Module

Protocols and training for National Healthcare Safety Network and the collection of CDI incidence data for reporting and feedback, from CDC

CDI Cause Analysis

Worksheet for reviewing CDI cases and adherence to facility policies and practices, including antibiotic review log, from the Massachusetts Coalition for the Prevention of Medical Errors

A Coordinated Response

A coordinated response is more effective than independent efforts¹

Facilities work together to protect patients.

Partners in Prevention:

The state and local health department and other state partners can assist and provide opportunities to extend efforts across the continuum of care

Common Approach (Not enough)

 Patients can be transferred back and forth from facilities for treatment without all the communication and necessary infection control actions in placa.

Independent Efforts (Still not enough

- Some facilities work independently to enhance infection control but are not often alerted to antibiotic-resistant or *C. difficile* germs coming from other facilities or outbreaks in the area.
- Lack of shared information from other facilities means that necessary infection control actions are not always taken and germs are spread to other patients.

Coordinated Approach (Needed)

- Public health departments track and alort health care facilities to antibioticresistant or *C. difficile* germs coming from other facilities and outbreaks in the area.
- Facilities and public health authorities share information and implement shared infection control actions to stop spread of germs from facility to facility.



Figure from CDC Vitals Signs: http://www.cdc.gov/vitalsigns/stop-spread/index.html

2007 Guideline for isolation precautions: preventing transmission of infectious agents in healthcare settings. Centers for Disease Control and Prevention Web site. <u>http://www.cdc.gov/hicpac/2007IP/2007isolation Published 2007</u>. Updated December 9, 2010. Accessed February 24, 2016.

Abad C., Fearday A., & Safdar N. (2010). Adverse effects of isolation in hospitalized patients. *J Hosp Infect, 76*, 97-102.

Ahyow L., Lambert P., Jenkins D., Neal K., & Tobin M. (2013). Occupancy rates and hospital-acquired *Clostridium difficile* infection: A cohort study. *Infect Control Hosp Epidemiol*, *34*, 1062-1069.

A practical guidance document for the laboratory detection of toxigenic *Clostridium difficile* infection. American Society of Microbiology Web site. <u>http://www.asm.org/images/pdf/Clinical/clostridiumdifficile9-21.pdf</u> Published September 21, 2010. Accessed February 24, 2016.

Berhe M., Edmond M., Bearman G. (2006). Measurement and feedback of infection control process measures in the intensive care unit: Impact on compliance. *Am J Infect Control*, *34*(8), 537-539.

Bliss D.Z., Johnson S., Savik L., Clabots C.R., Willard K., Gerding D.N. (1998). Acquisition of *Clostridium difficile* and *Clostridium difficile* associated disease in hospitalized patients receiving tube feeding. *Ann Intern Med*, *129*, 1011-1019.

Bobulsky G.S., Al-Nassir W.N., Riggs M.M., Sethi A.K., & Donskey C.J. (2008). *Clostridium difficile* skin contamination in patients with *C. difficile*-associated disease. *Clin Infect Dis*, 46(3), 447-450.

Boyce J.M., Havill N.L., Havill H.L., Mangione E., Dumigan D.G., Moore B.A. (2011). Comparison of fluorescent marker systems with 2 quantitative methods of assessing terminal cleaning practices. *Infect Control Hosp Epidemiol*, 32(12), 1187-1193.

Boyce J.M., Ligi C., Kohan C., Dumigan D., Havill N.L. (2006). Lack of association between the increased incidence of *Clostridium difficile*-associated disease and the increasing use of alcohol-based hand rubs. *Infect Control Hosp Epidemiol*, 27(5), 479-483.

Buendgens L., Bruensing J., Matthes M., et al. (2014). Administration of proton pump inhibitors in critically ill patients is associated with increased risk of developing *Clostridium difficile*-associated diarrhea. *J Crit Care, 696*, e11-15.

Carling P. (2013). Methods for assessing the adequacy of practice and improving room disinfection. *Am J Infect Control*, 41, S20-25.

Carrico, R. M., Rebmann, T., English, J. F., Mackey, J. Cronin, S. N. (2008). Infection prevention and control competencies for hospital based healthcare personnel. *Am J Infect Control*, 36; 691-701.

Chang H.T., Krezolek D., Johnson S., Parada J.P., Evans C.T., Gerding D.N. (2007). Onset of symptoms and time to diagnosis of *Clostridium difficile*-associated disease following discharge from an acute care hospital. *Infect Control Hosp Epidemiol*, 28(8), 926-931.

Cimiotti J.P., Aiken L.H., Sloane D.M., Wu E.S. (2012). Nurse staffing burnout, and health care associated infection. *Am J Infect Control, 40(6), 486-490*.

Dancer S.J., Kirkpatrick P., Corcoran D.S., Christison F., Farmer D., Robertson C. (2013). Approaching zero: temporal effects of a restrictive antibiotic policy on hospital-acquired *Clostridium difficile*, extended-spectrum β-lactamase-producing coliforms and methicillin-resistant *Staphylococcus* aureus. Int J Antimicrob Agents, 41(2), 137-142.

DeBono, S., Heling, G., Borg, M. A. (2014). Organizational culture and its implications for infection prevention and control in healthcare settings. *J Hosp Infect, 86*; 1-6.

Dubberke E.R., Carling P., Carrico, R., et al. (2014). Strategies to Prevent *Clostridium difficile* Infections in Acute Care Hospitals: 2014 Update. *Infect Control Hosp Epidemiol*, 35(6), 628-645.

Dubberke E.R. & Olsen M.A. (2012). Burden of *Clostridium difficile* on the Healthcare System. *Clin Infect Dis*, 55, S88-92.

Dubberke E.R., Reske K.A., Noble-Wang J., et al. (2007). Prevalence of *Clostridium dfficile* environmental contamination and strain variability in multiple health care facilities. *Am J Infect Control, 35*, 315-318.

Gonzalez E., Nandy P., Lucas A.D., Hitchins V.M. (2015). Ability of cleaning-disinfecting wipes to remove bacteria from medical device surfaces. *Am J Infect Control, 43*, 1331-1335.

Gould C.V., Edwards J.R., Cohen J., et al. (2013). Effect of nucleic acid amplification testing on populationbased incidence rates of *Clostridium difficile* infection. *Clin Infect Dis*, , 57, 1304-1307.

Griffiths P., Renz A., Hughes J., Rafferty A.M. (2009). Impact of organization and management factors on infection control in hospitals: a scoping review. *J Hosp Epidemiol*, 73, 1-14.

Gudnadottir U., Fritz J., Zerbel S. (2013). Reducing healthcare associated infections: Patients want to be engaged and learn about infection prevention. *Am J Infect Control*, *41*(*11*), 955-958.

Guidance to providers: Testing for C. difficile infection. Vanderbilt University Medical Center Web site. <u>http://www.mc.vanderbilt.edu/documents/infectioncontrol/files/Guidance%20for%20Providers%20</u> <u>FINAL%202011.pdf</u> Published August 31, 2011. Accessed February 24, 2016.

Guideline for disinfection and sterilization in healthcare facilities, 2008. Centers for Disease Control and Prevention Web site. <u>http://www.cdc.gov/hicpac/Disinfection_Sterilization/acknowledg.html</u>. Updated December 29, 2009. Accessed February 24, 2016.

Guidelines for Environmental Infection Control in Health Care Facilities, 2003. Centers for Disease Control and Prevention Web site. <u>http://www.cdc.gov/hicpac/pubs.html</u> Updated May 1, 2015. Accessed February 24, 2016.

HCUP Projections: Clostridium difficile Hospitalizations 2001 to 2013. HCUP Web site. <u>http://hcup-us.ahrq.gov/reports/projections/2014-01.pdf</u>. Published April 9, 2014. Accessed February 24, 2016.

Hensgens M.P., Goorhuis A., Dekkers O.M., Kuiiper E.J. (2012). Time interval of increased risk forClostridiumdifficile infection after exposure to antibiotics. J AntimicrobChemother, 67(3), 742-748.

Howell R., Novack V., Grgurich P., Souilliard D., Novack L., Pencina M., Talmor D. (2010). latrogenic gastric acid suppression and the risk of nosocomial *Clostridium difficile* infection. *Arch Int Med*, 170(9), 784-790.

Hsu J., Abad C., Dinh M., Safdar N. (2010). Prevention of endemic healthcare-associated Clostridium difficile infection: reviewing the evidence. *Am J Gastroenterol*, *105(11)*, 2327-2339.

Impact of Antibiotic Stewardship Programs on *Clostridium difficile (C. diff)* infections. Centers for Disease Control and Prevention Web site. http://www.cdc.gov/getsmart/healthcare/evidence/asp-int-cdiff.html Updated May 13, 2015. Accessed February 24, 2016.

Isaacson D., Haller B., Leslie H., Roemer M., Winston L. (2015). Novel handwashes are superior to soap and water in removal of *Clostridium difficile* spores from the hands. *Am J Infect Control*, 43, 530-532.

Johnson S., Gerding D.N., Olson M.M., Weiler M.D., Clabots C.R., Peterson L.R. (1990). Prospective, controlled study of vinyl glove use to interrupt *Clostridium difficile* nosocomial transmission. *Am J Med*, 88(2), 137-140.

Kombuj M., Sheahan A., Sun J., et al. (2016) Transmission of *Clostridium Difficile* during hospitalization for allogenic stem cell transplant. *Infect Control Hosp Epidemiol*, 37, 8-15.

Kufelnicka, A. M., Kim, T. J. (2011). Effective utilization of evolving methods for laboratory diagnosis of Clostridium difficile infection. *Clin Infect Dis*, 52; 1451-1452.

Kundrapu, S., Sunkesula, V., Jury, I., Deshpande, A., Donskey, C. (2014). A randomized trial of soap and water hand wash versus alcohol hand rub for removal of Clostridium difficile spores from hands of patients. *Infect Control Hosp Epidemiol*, 35; 204-205.

Lessa F., Mu Y., Bamberg W., Beldavs Z.G., et al. (2015). Burden of *Clostridium difficile* Infection in the United States. *N Engl J Med*, 372 (9), 825-834.

Linney S., Fernandes T., Einarson T., Sengar A. Walker J.H., Mills A. (2010). Association between use of proton pump inhibitors and a *Clostridium difficile*-associated disease outbreak: case-control study. *The Canadian Journal of Hospital Pharmacy*, 63(1), 31-37.

Litvin M., Reske K.A., Mayfield J., et al. (2009). Identification of a pseudo-outbreak of *Clostridium difficile* Infection (CDI) and the effect of repeated testing, sensitivity, and specificity on perceived prevalence of CDI. *Infect Control Hosp Epidemiol*, 30, 1166-1171.

Magill S.S, Edwards J.R., Bamberg W., et al. Emerging Infections Program Healthcare-Associated Infections and Antimicrobial Use Prevalence Survey Team. (2014). Multi State Point-Prevalence Survey of Health Care Associated Infections. *N Eng J Med*, 370, 1198-1208.

Mayfield J.L., Leet T., Miller J., Mundy L.M. (2003). Environmental control to reduce transmission of *Clostridium difficile*. *Clin Infect Dis*, 31(4), 995–1000.

McDonald L.C., Killgore G.E., Thompson A., et al. (2005). An epidemic, toxin gene–variant strain of *Clostridium difficile*. *N Eng JMed*, 353(23), 2433-2441.

McGukin M. & Govednik J. (2013) Patient empowerment and hand hygiene. J Hosp Infect, 84, 191-199.

Moore G., Smyth D., Singleton J., Wilson P. (2010). The use of adenosine triphosphate bioluminescence to assess the efficacy of a modified cleaning program implemented within an intensive care setting. *Am J Infect Control*, 38, 617-622.

Mulvey D., Redding P., Robertson C., Woodall C., Kingsmore P., Bedwell D., Dancer S.J. (2011). Finding a benchmark for monitoring hospital cleanliness. *J Hosp Infect*, *77*, 25-30.

Options for evaluating environmental cleaning. Centers for Disease Control and Prevention Web site. <u>http://www.cdc.gov/hai/toolkits/Evaluating-Environmental-Cleaning.html</u>. Published December 2010. Updated August 14, 2014. Accessed February 24, 2016.

Orenstein, R., Aronhalt, K., McManus, J. E. Fedraw, L. A. (2011). A targeted strategy to wipe out *Clostridium difficile. Infect Control Hosp Epidemiol, 32;* 1137-1139.

Oughton M.T., Loo V.G., Dendukuri N., Fenn S., Libman M.D. (2009). Hand hygiene with soap and water is superior to alcohol rub and antiseptic wipes for removal of *Clostridium difficile*. *Infect Control Hosp Epidemiol*, 30(10), 939-944.

Pepin J., Saheb N.B., Coulombe M.A. et al. (2005). The emergence of fluoroquinolones as the predominant risk factor for *Clostridium difficile*-Associated diarrhea: A cohort study during an epidemic in Quebec. *Clin Infect Dis*, 41(9), 1254-1260.

Peterson L.R., Robicsek A. (2009). Does my patient have *Clostridium difficile* infection? *Ann Int Med*, 151(3), 176-179.

Pincock T., Bernstein P., Warthman S., Holst E. (2012). Bundling hand hygiene interventions and measurement to decrease healthcare associated infections. *Am J Infect Control*, 40, S18-27.

Planche T., Aghaizu A., Holliman R. et al. (2008). Diagnosis of *Clostricium difficle* infection by toxin detection: a systematic review. *Lancet Infect Dis*, 8, 777-784.

Riggs M.M., Sethi A.K., Zabarsky T.F., Eckstein E.C., Jump R.L., Donskey C.J. (2007). Asymptomatic carriers are a potential source for transmission of epidemic and non-epidemic *Clostridium difficile* strains among long-term care facility residents. *Clin Infect Dis*, 45(8), 992–998.

Rutala W.A., Gergen M.F., Weber, D.J. (2012). Efficacy of different cleaning and disinfection methods against *Clostridium difficile* spores: importance of physical removal versus sporicidal inactivation. *Infect Control Hosp Epidemiol*, 33(12), 1255-1258.

See I., Mu Y., Cohen J., Beldavs Z.G. et al. (2014). NAP1 strain type *difficile* Infection. *Clin Infect Dis*, 58(10), 1394-1400. predicts outcomes from *Clostridium*

Sethi A.K., Al-Nassir W.N., Nerandzic M.M., Bobulsky G.S., Donskey C.J. (2010). Persistence of skin contamination and environmental shedding of *Clostridium difficile* during and after treatment of C. difficile infection. *Infect Control Hosp Epidemiol*, *31*(1), 21-27.

Shama G. & Malik D.J. (2013). Uses and abuses of rapid bioluminescence based ATP assays. *Int J Hyg Environ Health*, *216*, 1115-1125.

Shaughnessy M.K., Micielli R.L., DePestel D.D. Arndt J., Strachan C.L., Welch K.B., Chenoweth C.E. (2011). Evaluation of hospital room assignment and acquisition of *Clostridum difficile* infection *Infect Control Hosp Epidemiol*, 32, 201-206.

Sinkowitz-Cochran R.L., Burkitt K.H., Cuerdon T., et al. (2012). The associations between organizational culture and knowledge, attitudes, and practices in a multicenter Veterans Affairs quality improvement initiative to prevent methicillin-resistant *Staphylococcus aureus*. *Am J Infect Control*, 40, 138-143.

Sinkowitz-Cochran R.L., Garcia-Williams A., Hackbarth A.D., et al. (2012). Evaluation of organizational culture among different levels of healthcare staff participating in the Institute for Healthcare Improvement's 100,000 Lives Campaign. *Infect Control Hosp Epidemiol*, 33, 135-143.

Sitzlar, B., Deshpande, A., Fertelli, D., Kundrapu, S., Sethi, A., Donskey, C. (2013). An environmental disinfection odyssey: Evaluating of sequential interventions to improve disinfection of *Clostridium difficile* isolation rooms. *Infect Control Hosp Epidemiol*, 34; 459-465.

Slayton R.B., Toth D., Lee B.Y. et al. (2015). Vital Signs: Estimated Effects of a Coordinated Approach for Action to Reduce Antibiotic-Resistant Infections in Health Care Facilities - United States. *Morb Mort Week Rep*, 64 (30), 826-831.

Stabler R.A., Dawson L.F., Phua L.T., Wren B.W. (2008). Comparative analysis of BI/NAP1/027 hypervirulent strains reveals novel toxin B-encoding gene (tcdB) sequences. *J Med Micro*, 57(6), 771–775.

Stevens, V., Dumyati, G., Brown, J., van Wijngaarden, E. (2011). Differential risk of *Clostridium difficile* infection with proton pump inhibitor use by level of antibiotic exposure. *Pharmacoepidemiologic Drug Safety*, *20*; 1035-1042.

Sunenshine R.H. & McDonald L.C. (2006). *Clostridium difficile*-associated disease: new challenges from an established pathogen. *Cleve Clin J Med*, 73(2), 187-197.

Valiquette L., Cossette B., Garant M., Diab H., Pepin J. (2007). Impact of a reduction in the use of high-risk antibiotics on the course of an epidemic of *Clostridium difficile* associated disease caused by the hypervirulent ANP1/027 strain. *Clin Infect Dis*, 45, S112-121.

 Warny M., Pepin J., Fang A., et al. (2005). Toxin production by and emerging strain of *Clostridium difficile* associated with outbreaks of severe disease in North America and Europe. *The Lancet*, 366(9491), 1079-1084.

Weber D. & Rutala W. (2011). The role of the environment in transmission of *Clostridium difficile* transmission in healthcare facilities. *Infect Control Hosp Epidemiol*, 32, 207-209.

Wilcox M.H., Fawley W.N., Wigglesworth N., Parnell P., Verity P., Freeman J. (2003). Comparison of the effect of detergent versus hypochlorite cleaning on environmental contamination and incidence of *Clostridium difficile* infection. *J Hosp Infect*, 54(2), 109-114.